

This article was downloaded by:

On: 29 January 2011

Access details: *Access Details: Free Access*

Publisher *Taylor & Francis*

Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



## Phosphorus, Sulfur, and Silicon and the Related Elements

Publication details, including instructions for authors and subscription information:

<http://www.informaworld.com/smpp/title~content=t713618290>

### Synthetic Analogues of the Second Messenger D-MYO Inositol 1,4,5 Trisphosphate as Receptor Agonists and Inhibitors of the Enzymes of the Polyphosphoinositide Pathway of Signal Transduction

Barry V. L. Potter<sup>a</sup>

<sup>a</sup> School of Pharmacy and Pharmacology, University of Bath, Bath, U.K.

**To cite this Article** Potter, Barry V. L.(1993) 'Synthetic Analogues of the Second Messenger D-MYO Inositol 1,4,5 Trisphosphate as Receptor Agonists and Inhibitors of the Enzymes of the Polyphosphoinositide Pathway of Signal Transduction', *Phosphorus, Sulfur, and Silicon and the Related Elements*, 76: 1, 143 — 146

**To link to this Article:** DOI: 10.1080/10426509308032379

**URL:** <http://dx.doi.org/10.1080/10426509308032379>

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: <http://www.informaworld.com/terms-and-conditions-of-access.pdf>

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

## SYNTHETIC ANALOGUES OF THE SECOND MESSENGER D-MYO INOSITOL 1,4,5 TRISPHOSPHATE AS RECEPTOR AGONISTS AND INHIBITORS OF THE ENZYMES OF THE POLYPHOSPHOINOSITIDE PATHWAY OF SIGNAL TRANSDUCTION

BARRY V L POTTER

School of Pharmacy and Pharmacology, University of Bath, Claverton Down, Bath BA2 7AY, U.K.

**Abstract** Novel synthetic probes of the polyphosphoinositide pathway of signal transduction are discussed.

### INTRODUCTION

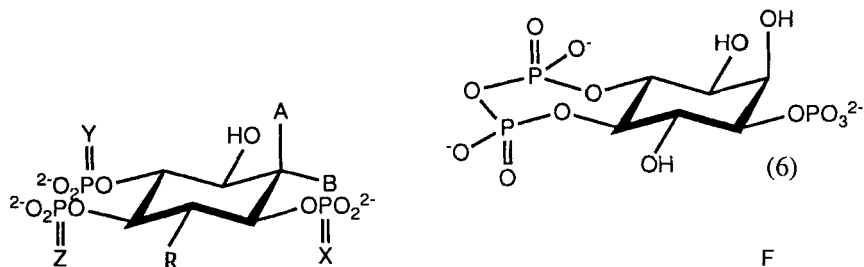
D-*myo*-inositol 1,4,5-trisphosphate [ $\text{Ins}(1,4,5)\text{P}_3$ ] **1** is a ubiquitous intracellular messenger that couples stimulation of cell surface receptors to the release of intracellular  $\text{Ca}^{2+}$ . An  $\text{Ins}(1,4,5)\text{P}_3$  receptor has been purified<sup>2</sup>, reconstituted<sup>3</sup>, an amino acid sequence determined from the cDNA<sup>4,5</sup> and a sequence near to the N-terminus is responsible for  $\text{Ins}(1,4,5)\text{P}_3$  binding<sup>6</sup>. Because of the fundamental importance of the polyphosphoinositide signalling system it is desirable to have access to pharmacological tools for intervention at  $\text{Ins}(1,4,5)\text{P}_3$  receptors and inhibition of the metabolic enzymes acting upon this second messenger<sup>7,8</sup>.  $\text{Ins}(1,4,5)\text{P}_3$  antagonists and enzyme inhibitors may have therapeutic potential<sup>9</sup>. Few such tools have yet been identified and there are significant difficulties intrinsic to a drug design strategy based upon  $\text{Ins}(1,4,5)\text{P}_3$ .

Synthetic approaches to inositol polyphosphates addressed the problems of selective protection, phosphorylation, resolution and deblocking<sup>10,11</sup>. The design and synthesis of almost any inositol polyphosphate analogue can now be envisaged<sup>10-12</sup>. The first example was *myo*-inositol 1,4,5-trisphosphorothioate [ $\text{Ins}(1,4,5)\text{PS}_3$ ] **2**<sup>13</sup>. Further analogues have since been synthesized, i.e. inositol 1,4-bisphosphate-5-phosphorothioate **3**<sup>14</sup>, inositol 1-phosphorothioate-4,5-bisphosphate **4**<sup>15</sup> and inositol 1-phosphate-4,5-bisphosphorothioate **5**<sup>16</sup>. Their resistance to enzymic degradation<sup>17</sup> makes them valuable pharmacological tools. We focus here on inositol phosphorothioates and other novel agonists and enzyme inhibitors synthesized by our group. We have also recently synthesised an interesting pyrophosphate analogue of  $\text{Ins}(1,4,5)\text{P}_3$ , **6**<sup>16</sup>.

Numerous synthetic routes to  $\text{Ins}(1,4,5)\text{P}_3$  have now been devised<sup>10-12</sup>. Our initial route<sup>18</sup> exploited methodology used to synthesize *myo*-inositol 4,5-bisphosphate and involved phosphorylation of the precursor, L-1,2,4-tri-*O*-benzyl-*myo*-inositol<sup>19</sup>. After deblocking and purification, the resulting  $\text{Ins}(1,4,5)\text{P}_3$  was fully active at mobilising  $\text{Ca}^{2+}$  and binding to the cerebellar  $\text{Ins}(1,4,5)\text{P}_3$  receptor.  $\text{Ins}(1,4,5)\text{PS}_3$  Was synthesized by a modification of the procedure<sup>13</sup>.  $\text{Ins}(1,4,5)\text{PS}_3$  binds with high affinity to  $\text{Ins}(1,4,5)\text{P}_3$  receptor sites in brain<sup>17,20</sup> and liver<sup>21</sup>. It is a full  $\text{Ca}^{2+}$ -releasing agonist, being only some 3-4 fold less potent than  $\text{Ins}(1,4,5)\text{P}_3$  in a variety of cells, such as in *Xenopus* oocytes<sup>22,23</sup>, permeabilised Swiss 3T3

cells<sup>22,24</sup>, GH<sub>3</sub> cells<sup>24</sup>, hepatocytes<sup>25</sup>, pancreatic<sup>26-28</sup> and parotid<sup>29</sup> acinar cells, skeletal muscle triads<sup>30</sup>, mouse lacrimal cells<sup>31</sup> and SH-SY5Y cells<sup>7</sup>. It is resistant to dephosphorylation by 5-phosphatase<sup>17,25</sup> and can produce a sustained cellular calcium transient<sup>7,25</sup>. Ins(1,4,5)PS<sub>3</sub> is a potent competitive inhibitor of 5-phosphatase<sup>32</sup>, but does not compete with Ins(1,4,5)P<sub>3</sub> for 3-kinase<sup>25-33</sup>. Applications of inositol phosphorothioates have been reviewed<sup>34</sup>.

Applications of inositol phosphorothioates detailed above have been extended by the commercial availability of the metabolically stable radioligand, [<sup>35</sup>S]-Ins(1,4,5)PS<sub>3</sub>. [<sup>35</sup>S]-D-Ins(1,4,5)PS<sub>3</sub> was prepared by a modification to the chemical synthesis of unlabelled material<sup>13</sup>. Characterisation of its interaction with cerebellar membranes shows that D-[<sup>35</sup>S]-Ins(1,4,5)PS<sub>3</sub> binds with high affinity to the Ins(1,4,5)P<sub>3</sub> receptor<sup>35</sup>, labels more sites than does [<sup>3</sup>H]-Ins(1,4,5)P<sub>3</sub>, and may label different conformations of the receptor with equal affinity, whereas Ins(1,4,5)P<sub>3</sub> induces or stabilises some of the sites in a high affinity form.



A=OH; B=H; R=OH; X=Y=Z=O (1)

X=Y=Z=S (2)

X=Y=O; Z=S (3)

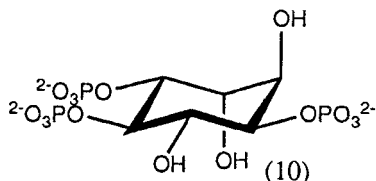
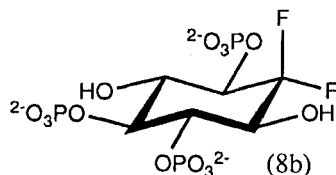
X=S; Y=Z=O (4)

X=O; Y=Z=S (5)

A=H; B=F; R=OH; X=Y=Z=O (7ab)

A=B=F; R=OH; X=Y=Z=O (8a)

A=OH; B=H; R=H; X=Y=Z=O (9)



(a) **Fluorinated analogues 8ab** Inositol ring-modified and phosphate-modified analogues have been synthesized<sup>10,11</sup> to understand the role of recognition motifs of Ins(1,4,5)P<sub>3</sub> in receptor binding, specificity and stimulation. We synthesised 2-deoxy-2-fluoro-*scyllo* Ins(1,4,5)P<sub>3</sub> 7ab and 2,2-difluoro-2-deoxy Ins(1,4,5)P<sub>3</sub> 8ab and studied their interaction with the Ca<sup>2+</sup>-releasing receptor and Ins(1,4,5)P<sub>3</sub> 5-phosphatase and 3-kinase<sup>36</sup>. Unexpectedly, DL-(8ab), was not a substrate for 5-phosphatase. We suspected that L-(8b) might act as a 5-phosphatase inhibitor and we resolved DL-(8ab) into its enantiomers<sup>37</sup> and studied their differential interactions with the three Ins(1,4,5)P<sub>3</sub> binding proteins<sup>38</sup>. D-(8a) was a full agonist and released Ca<sup>2+</sup> with kinetics similar to Ins(1,4,5)P<sub>3</sub> and with an EC<sub>50</sub> of 0.21 μM. Fluoro-substitution at the 2-position therefore makes D-(8a) an excellent analogue of Ins(1,4,5)P<sub>3</sub> and essentially equipotent to it. D-(8a) was a substrate for Ins(1,4,5)P<sub>3</sub> 3-kinase and inhibited Ins(1,4,5)P<sub>3</sub> phosphorylation potently [apparent K<sub>i</sub>, 10.2 μM]. L-(8b) inhibited phosphorylation of Ins(1,4,5)P<sub>3</sub> [K<sub>i</sub>, 11.9 μM] and is

a potent 3-kinase inhibitor. 5-Phosphatase is relatively non-selective for inositol phosphates, but 3-kinase is very specific<sup>7</sup>, posing problems in inhibitor design. L-(8b) represents a lead in the search for a small molecule 3-kinase inhibitor.

Ins(1,4,5)P<sub>3</sub> and D-(8a) were comparable substrates for 5-phosphatase but treatment of L-(8b) with 5-phosphatase did not liberate inorganic phosphate and racemic DL-(8ab) was apparently not a substrate<sup>36</sup> but an inhibitor with a K<sub>i</sub> of 19 μM. Fluorination at the 2-position of L-Ins(1,4,5)P<sub>3</sub> has led to the production of a small molecule 3-kinase inhibitor and a moderately potent 5-phosphatase inhibitor which is a very poor agonist for Ca<sup>2+</sup> release.

(b) 6-Deoxy-*myo*-inositol 1,4,5-trisphosphate 9 A key structure-activity aspect concerns the potential role of the 6-OH of Ins(1,4,5)P<sub>3</sub>, adjacent to the crucial vicinal 4,5-bisphosphate, in determining the affinity and specificity of Ins(1,4,5)P<sub>3</sub> for its receptor and the metabolic enzymes 3-kinase and 5-phosphatase. We have thus deleted the 6-OH of Ins(1,4,5)P<sub>3</sub> to generate 6-deoxy Ins(1,4,5)P<sub>3</sub> **9**<sup>39</sup>. **9** has an EC<sub>50</sub> for Ca<sup>2+</sup> release of 6.4 μM and is a full agonist. Deletion of the 6-OH makes **9** approximately 70-fold less potent than Ins(1,4,5)P<sub>3</sub>, suggesting a lower affinity for the receptor<sup>39</sup>. **9** Inhibited 5-phosphatase [K<sub>i</sub>, 76 μM] suggesting that the 6-OH may play a key role in the catalytic mechanism of this enzyme.

(c) L-Chiro-inositol 2,3,5-trisphosphate 10 Generation of a 3-kinase inhibitor could arise by modification of the 3-OH of Ins(1,4,5)P<sub>3</sub>. We developed a concise synthetic route to L-Chiro-inositol 2,3,5-trisphosphate **10**, i.e. Ins(1,4,5)P<sub>3</sub> with an inverted 3-OH group, from L-quebrachitol<sup>40</sup>. **10** Was a potent agonist for the release of intracellular Ca<sup>2+</sup> with an EC<sub>50</sub> some 5-10 fold higher than Ins(1,4,5)P<sub>3</sub>. It was not a substrate for 3-kinase (K<sub>i</sub>, 7.1 μM), nor surprisingly for 5-phosphatase (K<sub>i</sub>, 7.7 μM), and released Ca<sup>2+</sup> in a sustained fashion similar to Ins(1,4,5)P<sub>3</sub>.

We have already noted that deletion of the 6-OH of Ins(1,4,5)P<sub>3</sub> generates a moderately potent 5-phosphatase inhibitor in **9**<sup>39</sup> of this enzyme. However, the finding that L-chiro-Ins(2,3,5)P<sub>3</sub> is a potent inhibitor was unexpected and poses the question as to how a single change in orientation of a remote OH group can have such a radical effect. Whether this is the result of subtle conformational changes induced in the molecule relative to Ins(1,4,5)P<sub>3</sub>, or of non-productive binding of **10** to the enzyme present intriguing prospects for further investigation.

**ACKNOWLEDGEMENTS:** Work reported here was supported by SERC (MRI) and The Wellcome Trust. B.V.L.P. is a Lister Institute Fellow.

## REFERENCES

1. M.J. Berridge, *Annu. Rev. Biochem.*, **56**, 159 (1987).
2. S. Supattapone, P.F. Worley *et al*, *J. Biol. Chem.*, **263**, 1530 (1988).
3. C.D. Ferris, R.L. Haganir, S. Supattapone *et al*, *Nature*, **342**, 87 (1989).
4. T. Furuichi, S. Yoshikawa, A. Miyawaki *et al*, *Nature*, **342**, 32 (1989).
5. G.A. Mignery, T.C. Südhof, K. Takei *et al*, *Nature*, **342**, 192 (1989).
6. G.A. Mignery, C.L. Newton *et al*, *J. Biol. Chem.*, **265**, 12679 (1990).
7. S.R. Nahorski and B.V.L. Potter, *Trends Pharmacol. Sci.*, **10**, 139 (1989).
8. T.K. Ghosh, P.S. Eis, J.M. Mullaney *et al*, *J. Biol. Chem.*, **263**, 11075 (1988).
9. E.R. Chilvers, E.D. Kennedy and B.V.L. Potter, *Drug News and Perspectives*, **2**, 342 (1989).
10. D.C. Billington, *Chem Soc. Rev.*, **18**, 83 (1989).

11. B.V.L. Potter, Nat. Prod. Reports, **7**, 1 (1990).
12. B.V.L. Potter, in Transmembrane Signalling, Intracellular Messengers and Implications for Drug Development, edited by S.R. Nahorski (Wiley, Chichester, U.K., 1990).
13. A.M. Cooke, R. Gigg and B.V.L. Potter, J. Chem. Soc. Chem. Commun., 1525 (1987).
14. A.M. Cooke, N.J. Noble, R. Gigg, S. Payne and B.V.L. Potter, J. Chem. Soc. Chem. Commun., 269 (1988).
15. D. Lampe and B.V.L. Potter, ibid., 1500 (1990).
16. N.J. Noble and B.V.L. Potter, Bioorg. Med. Chem. Lett., **2**, 471 (1992).
17. A.L. Willcocks, B.V.L. Potter, A.M. Cooke and S.R. Nahorski, Eur. J. Pharmacol., **155**, 181 (1988).
18. A.M. Cooke, R. Gigg and B.V.L. Potter, Tet. Lett., **28**, 2305 (1987).
19. J. Gigg, R. Gigg *et al.*, J. Chem. Soc. Perkin Trans. I., 423 (1987).
20. A.L. Willcocks, A.M. Cooke, B.V.L. Potter and S.R. Nahorski, Biochem. Biophys. Res. Commun., **146**, 1071 (1987).
21. D.L. Nunn, B.V.L. Potter and C.W. Taylor, Biochem. J., **265**, 393 (1990).
22. C.W. Taylor, M.J. Berridge, K.D. Brown, A.M. Cooke and B.V.L. Potter, Biochem. J., **150**, 626 (1988).
23. S. DeLisle, K.-H. Krause, G. Denning, B.V.L. Potter and M.J. Welsh, J. Biol. Chem., **265**, 11726 (1990).
24. J. Strupish, A.M. Cooke, B.V.L. Potter, R. Gigg and S.R. Nahorski, Biochem. J., **253**, 901 (1988).
25. C.W. Taylor, M.J. Berridge, A.M. Cooke and B.V.L. Potter, Biochem. J., **259**, 645 (1989).
26. F. Thevenod, M. Dehlinger-Kremer, T.P. Kemmer, A.-L. Christian, B.V.L. Potter and I. Schulz, J. Membr. Biol., **109**, 173 (1989).
27. M. Wakui, B.V.L. Potter and O.H. Petersen, Nature, **339**, 317 (1989).
28. C.C.H. Petersen, E.C. Toescu, B.V.L. Potter and O.H. Petersen, FEBS Lett., **293**, 179 (1991).
29. F.S. Mennitti, H. Takemura, O. Thastrup, B.V.L. Potter and J.W. Putney Jr., J. Biol. Chem., **246**, 13646 (1991).
30. C. Valdivia, H.H. Valdivia, B.V.L. Potter and R. Coronado, Biophys. J., **57**, 1233 (1990).
31. L. Changya, D.V. Gallacher, R.F. Irvine, B.V.L. Potter and O.H. Petersen, J. Membr. Biol., **109**, 85 (1989).
32. A.M. Cooke, S.R. Nahorski and B.V.L. Potter, FEBS Lett., **242**, 373 (1989).
33. S.T. Safrany, R.J.H. Wojcikiewicz, J. Strupish, J. McBain, A.M. Cooke, B.V.L. Potter and S.R. Nahorski, Mol. Pharmacol., **39**, 754 (1991).
34. B.V.L. Potter and S.R. Nahorski, Biochem. Soc. Trans., **20**, 434 (1992).
35. R.A.J. Challiss, S.M. Smith, B.V.L. Potter, B.V.L. and S.R. Nahorski, FEBS Lett., **281**, 101 (1991).
36. S.T. Safrany, D. Sawyer, R.J.H. Wojcikiewicz, S.R. Nahorski and B.V.L. Potter, FEBS Lett., **276**, 91 (1990).
37. D. Sawyer and B.V.L. Potter, Bioorg. Med. Chem. Lett., **1**, 705 (1991).
38. S.T. Safrany, D. Sawyer, S.R. Nahorski and B.V.L. Potter, Chirality (1992) in press.
39. S.T. Safrany, R.J.H. Wojcikiewicz, J. Strupish, D. Dubreuil, J. Cleophax, S.D. Gero, S.R. Nahorski and B.V.L. Potter, FEBS Lett., **278**, 252 (1991).
40. C. Liu, S.R. Nahorski and B.V.L. Potter, J. Chem. Soc. Chem. Commun., 1014 (1991).